

FACS ARRAY USER GUIDE

General: NO FITC!!!

Colors that can be used:

PerCP-Cy5.5 or PE-Cy7

PE, APC-Cy7, APC

Laser	Fluorochromes		Detector Parameter
532 nm	PE		Yellow
	PE-Cy7	PerCP-Cy5.5	Far Red
635 nm	APC	Alexa Fluor 647	Red
	APC-Cy7		Near Infra Red (NIR)

FACS Sheath: special for FACS Array

Starting Up procedure

1. Power Switch on the backside of the machine in the middle
2. Power Switch on the front side
3. Switch on the computer
 - Username: Administrator
 - Password: BDIS

Click on BD FACS Array System Software

Login name: Administrator

Password: welcome

BD FACS Array Software

Open *Experiment Wizard* and select a Wizard session:

- Screening clones
- CBA
- HLA typing
- Tetramer Titration

Next

Choose template: ? **colors I)** or **CBA II)**

Next



I) Normal FACS analysis

Follow the protocol of the Wizard step by step:

Instrument Setup and optical Spillover (automatic compensation) – **Yes**

Next

Click on the **colors** you will use

Next

Number of samples – settings not included !

Next

Sample identification – you can add a name

Next

Number of wells per sample

Next

Name of well

Next

- Fluorophore labels – **enter the name of the antibodies**
- Next
- Keyword
- Next
- Adjusting the plate layout – **continuous horizontal without separator**
- Next
- Loader Settings: **Sample flow rate 2.0 μ l/sec**
Sample volume: 60 μ l if you have 100 μ l in the well
 Mixing volume 50 μ l
 Mixing speed 200 μ l
 Number of mix 3
 Wash volume 200 μ l
- Next
- Acquisition settings – **how many events**
- Next
- Experiment name - **Initials (2 letters) followed by the date**
- Next
- Saving Wizard session – Yes, if you like to save the experiment as template in the Experiment Wizard
- Next
- A summary of your designed experiment appears on the screen. If you like to modify go <Back> - otherwise
- Finish**

Your Experience appears on the workspace



Setting layout on the plate:

unstained
 PerCP-Cy5.5/Pe-Cy7
 PE
 APC-Cy7
 APC

Open *Setup Tools* –
Instrument/Parameters to open compensation parameters




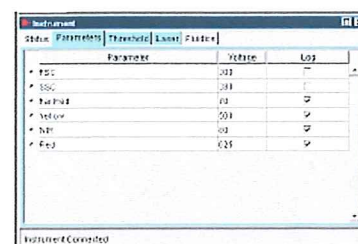
Load your plate (click load)

In the Workspace go on *plate_001/ Display: Acquire*

In *Select* click on **None** (wells will turn white)

Click in the Acquire Sheet on well **A1** (it turn green)

Click on Inspector  *Acquisition - Experiment* and change the flow rate to **0.5 μ l/sec** and the sample volume to **100 μ l** for the settings



Click on *Setup* and measure a **few** events – click on **setup** again to stop

Settings:

Click on **plate_001/ Acquire/ Select/ None**

Adjust the FSC and SSC voltage to place the population of interest on the scale in the plot. Adjust the voltage of each fluorochrome to ~300 (baseline) and press enter.

Click on the next well **A2: Setup**

- ❖ Adjust gate in dot plot to lymphocytes
- ❖ Adjust voltage of the fluorochrome if necessary
- ❖ Adjust Gate P2 to the peak if necessary

Repeat the procedure for each color

Save Settings to calculate the compensation

Go back to well **A1** click on **Acquire**, the samples will pass automatically and the compensation will be calculated

Make sure that you have enough sample volume in each well!!!!

Acquire your samples

Go in *Select* to **AUTO**

Click in *Acquisition* on **Acquire** – all programmed wells will be acquired and saved automatically.

If you have more than one plate unload the first, measured one and load the second plate and click on **acquired**.

Export data



Click on the Browser
Select Experiment

File/BDExport/LICR/Month folder

FCS – Check FCS format :2.0 for Cellquest; 3.0 for FACS Diva

II) CBA-Experiments

Open *Experiment Wizard*

Next

Choose template: **BD CBA Kit**

OKay

Follow the protocol of the Wizard step by step, - **P1 as acquisition stop!!**

NO SETTINGS AND NO CONTROLS ARE NEEDED !!

Export data in folder **Export>CBA**

Cleaning Procedure and Shut down

Go in Display/ Acquire and click on Unload

- 1) Use the **Clean** function in the *Instrument* menu
- or
- 2) Add the samples at the end your of your samples on the plate

Put in 96 well plate 4 wells FACS Clean followed by 4 wells dH₂O

Go to Display *Prepare* and add the samples manually by clicking on the first well after your last sample

Display: Acquire/plate_001/Load - **Acquire**

When finish: Display: Acquire/plate_001/**Unload**

Close your Experiment

Click on **Shutdown Fluidics** on the Instrument menu

Wait till the fluidics is shut down - message appears in the Status box – very fast!

Close the BD FACS Array System Software

Shut down the computer and switch of the **two** power buttons

Fill up the Sheath

Empty the Waste each time after use