Long-Term Soil Structure Observatory for Monitoring Post-Compaction Evolution of Soil Structure

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The projected intensification of agriculture to meet food targets of a rapidly growing world population are likely to accentuate already acute problems of soil compaction and deteriorating soil structure in many regions of the world. The key role of soil structure for soil functions, the sensitivity of soil structure to agronomic management practices, and the lack of reliable observations and metrics for soil structure recovery rates after compaction motivated the establishment of a long-term Soil Structure Observatory (SSO) at the Agroscope research institute in Zürich, Switzerland. The primary objective of the SSO is to provide long-term observation data on soil structure evolution after disturbance by compaction, enabling quantification of compaction recovery rates and times. The SSO was designed to provide information on recovery of compacted soil under different post-compaction soil management regimes, including natural recovery of bare and vegetated soil as well as recovery with and without soil tillage. This study focused on the design of the SSO and the characterization of the pre- and post-compaction state of the field. We deployed a monitoring network for continuous observation of soil state variables related to hydrologic and biophysical functions (soil water content, matric potential, temperature, soil air O₂ and CO₂ concentrations, O₂ diffusion rates, and redox states) as well as periodic sampling and in situ measurements of infiltration, mechanical impedance, soil porosity, gas and water transport properties, crop yields, earthworm populations, and plot-scale geophysical measurements. Besides enabling quantification of recovery rates of compacted soil, we expect that data provided by the SSO will help improve our general understanding of soil structure dynamics.

Soil compaction due to agricultural operations is a serious threat to soil productivity and soil ecological functions in modern agriculture (Hamza and Anderson, 2005; Schjønning et al., 2015). The susceptibility of soil to compaction is a combination of inherent soil properties, soil moisture status at the time of field operations, soil and land management, and the machinery used. Soil compaction problems are therefore especially widespread in agriculture involving heavy machinery and/or moist soils; such is the case in large parts of Europe and North America or parts of South America and Australia. According to Jones et al. (2003), more than one third of European subsoils are highly susceptible to compaction, and Schjønning et al. (2015) suggested that a quarter of all European soils were compacted. Hamza and Anderson (2005) cited figures of compacted areas of 68 Mha (worldwide), 33 Mha (Europe), and 4 Mha (Australian wheat belt). The aggravation of soil compaction is linked with modern trends of steadily

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increasing power and weight of agricultural vehicles and implements, which today may reach wheel loads in excess of 10 Mg for self-propelled harvesters or slurry spreaders (Schjønning et al., 2015). Economical and efficiency considerations suggest no slowing of this trend unless a technological breakthrough or a paradigm shift (e.g., small autonomous vehicles, fixed tracks) would occur. Considering the increasing pressures on agriculture for production of food, fiber, and biofuels, the continual degradation of soil resources due to compaction is expected to escalate in the future. Many modern agricultural vehicles induce stresses in the soil that exceed the mechanical strength of soils under most conditions, resulting in soil deformation and modification of the spatial arrangement of soil constituents and voids (poles), altering a critical component of soil structure. The reduction in pore volume and the change in pore structure (size distribution, connectivity, tortuosity) decrease transport capability (fluid and gas transport by advection and diffusion) and water storage capacity, increase mechanical resistance for root growth, change prevailing chemical reactions and the availability of nutrients (anoxic conditions), and modify physical habitats of soil organisms. These changes consequently affect biodiversity and a range of ecosystem services including the production of food, fiber, and fuel, water provision and purification, and atmospheric regulation (e.g., Zhang et al., 2007).

The real costs of soil compaction are borne by the cumulative loss of soil functionality (e.g., yield loss in agricultural systems) following a significant compaction event, integrated over the time period until a soil has effectively recovered to its pre-compaction functionality. Hence, quantifying compaction costs (and therefore the severity of compaction) requires knowledge about soil structure recovery rates after compaction. We differentiate between instantaneous compaction impact and compaction damage (Fig. 1), and define instantaneous compaction impact as the immediate effect of a compaction event on soil functions, whereas compaction damage relates to the compaction effect integrated over the structure recovery time. Considering the conceptual sketch in Fig. 1, we may describe compaction damage (CD) as

$$CD = \int \left[ CI \left(1 - C_r(t) \right) \right] dt$$

where CI is the immediate compaction impact (immediate change in a soil metric), $C_r(t)$ is the soil structure recovery rate (a function of time), and $t$ is the elapsed time since compaction. The compaction damage can be expressed in terms of compaction costs, i.e., by valuation of ecosystem services, although this is not an easy exercise (e.g., Loomis et al., 2000; Farber et al., 2002).

The costs of compaction relate to ecological costs but also to direct monetary value, e.g., for farmers in terms of reduced profit due to yield decline. A review of compaction costs was given by Chamen et al. (2015). We note that partial recovery times, e.g., 90% recovery, are typically used in ecological studies (e.g., MacNeil et al., 2015). Partial recovery times provide more useful estimates because simplistic recovery functions (e.g., logarithmic functions) may display unrealistic asymptotic behavior at large recovery times (Webb and Wilshire, 1980; Webb, 2002). While recovery describes a process, resilience is defined as the ability to recover a property, capacity, or function to its initial value when the disturbance (i.e., stress) is removed (e.g., Lal, 1993; Gregory et al., 2007). Resilience is typically quantified in terms of a resilience index that relates the initial impact (i.e., impact at time 0) to the effect at time $x$ (e.g., Orwin and Wardle, 2004). Hence, resilience quantifies the amount of recovery at a certain time step after the disturbance.

Our knowledge regarding soil compaction recovery rates and recovery times is anecdotal and sketchy. Experimental evidence of compacted soil recovery periods from field studies suggests a wide range, from months (Drewry et al., 2004) and years (Culley et al., 1982; Blackwell et al., 1985; Langmaack et al., 1999; Radford et al., 2007; Besson et al., 2013) to decades (Håkansson and Reeder, 1994; Peng and Horn, 2008; Berisso et al., 2012) and centuries (Webb, 2002). In these studies, only a few properties and functions were addressed. Traditionally, crop yield has been the focus of such evaluations (e.g., Culley et al., 1982; Håkansson and Reeder, 1994), while more recent work has addressed soil structure and transport functions (e.g., Peng and Horn, 2008; Berisso et al., 2012; Besson et al., 2013) or earthworm abundance (Langmaack et al., 1999). Estimates of recovery rates based on laboratory studies (e.g., Arthur et al., 2013) are typically higher than those observed in field studies, thus potentially underestimating recovery times and compaction costs. The conflicting evidence of compaction recovery times, and the discrepancy in recovery rate estimates between

![Fig. 1. Schematic figure showing immediate compaction impact, compaction recovery rate $C_r(t)$, compaction recovery time $t_r$, and compaction damage. Case 1 in blue indicates a large immediate impact but a fast recovery, thus moderate damage (e.g., topsoil compaction), while Case 2 (red) shows a moderate immediate impact but slow recovery, thus large damage (e.g., subsoil compaction).](image-url)
A more definitive understanding of compacted soil recovery rates and times is not only needed for estimation of the real costs of compaction (see above and Fig. 1), but a more quantitative description of natural compaction recovery mechanisms and recovery pathways could also be used to develop site-specific soil management methods (or strategies) that accelerate compaction recovery (Dexter, 1991). More generally, improved understanding of soil structure dynamics due to biophysical processes could offer a path for harnessing biological processes to improve the physical and ecological conditions of soil for agricultural production (Dexter, 1991; Hallett et al., 2013). Protocols for enhancing soil structure recovery (and more generally, soil structure formation) are urgently needed, with the growing threat of soil compaction as a result of the intensification of agriculture to enhance food production.

A framework for combining in situ monitoring, modeling, and small-scale mechanistic experiments targeting the different compaction recovery processes is necessary to better understand the complex biophysical interactions and feedback mechanisms involved in soil structure dynamics. A major gap in our knowledge of soil structure dynamics as identified above is the discrepancy in temporal and spatial scales between laboratory investigations and field studies under natural climatic conditions and the lack of systematic field observations at the proper (i.e., years to decades) time scale. Consequently, research on soil structure evolution requires, among others, long-term field studies including adequate research infrastructure for monitoring.

To address these challenges, we designed a long-term field experiment for monitoring post-compaction evolution of soil structure, referred to as a soil structure observatory. Here we describe the objectives, the design, the implementation, and monitoring concept of our soil structure observatory and present the initial compaction effects.
The site (~1.5 ha) was sown with a grass mixture in April 2013, roughly 1 yr before initiating soil compaction in March 2014. We used the grass–white clover (Trifolium repens L.) mixture SM 442 (Suter et al., 2008) reinforced with an additional 4 kg ha⁻¹ of lucerne (Medicago sativa L.). This grass mixture was chosen because it contains species with different root systems and depth-distributions of roots, and because it is known to be robust under a wide range of unfavorable soil physical conditions (e.g., water logging and O₂ stress). After sowing the mixture, no more traffic with agricultural machinery was allowed over the experimental area, except for a small self-propelled mower used for periodic cutting of the grass. The initiation of the ley 1 yr prior to initial compaction and the prohibition of any farm machinery traffic aimed at allowing the soil to “rest” before establishing the SSO.

A permanent meteorological station run by MeteoSwiss is located within 300 m from the experimental field, delivering hourly data on, among others, air temperature, precipitation, relative humidity, and global radiation.

Experimental Design

The SSO includes two experimental factors—compaction level and post-compaction soil management (see below)—and is based on a strip-plot design (Fig. 2) with three blocks (replicates). There are three compaction levels and four management treatments, as detailed below, amounting to a total of 36 plots. Each plot is 17 m long and 10 to 12 m wide. A one-time compaction was performed in the in-sown ley (see above) in March 2014, and the four post-compaction soil management treatments were immediately started thereafter. Thus, the (soil) conditions during compaction were the same (ley) in all treatments. No further experimental compaction is foreseen in the SSO.

Compaction Treatments

The SSO includes three compaction levels (Fig. 2): compaction of the entire plot area, i.e., track-by-track (CEP), compaction in wheel tracks (CWT), and control, i.e., no experimental compaction (NOC). The same compaction intensity (i.e., the same machinery and the same number of machinery passes) was applied in CEP and CWT.

Treatment CWT allows studying soil structure evolution under conditions where roots and soil organisms can avoid compacted zones (wheel tracks) (Capowiez et al., 2009), in comparison with processes under CEP, where the entire near-surface volume is compacted. Soil compaction recovery might be slower in the compacted zones under the wheel tracks if roots and soil organisms

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Table 1. Basic soil properties at the Soil Structure Observatory, Zürich, Switzerland.

<table>
<thead>
<tr>
<th>Block</th>
<th>Clay (&lt;2 μm)</th>
<th>Silt (2–50 μm)</th>
<th>Sand (50–2000 μm)</th>
<th>Organic C</th>
<th>Particle density</th>
<th>pH (CaCl₂)</th>
<th>Cation exchange capacity</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>g g⁻¹</td>
<td></td>
<td></td>
<td></td>
<td>Mg m⁻³</td>
<td></td>
<td>cmolₑ kg⁻¹</td>
</tr>
<tr>
<td>0–0.2-m depth</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>0.254 (0.014)†</td>
<td>0.518 (0.015)</td>
<td>0.228 (0.018)</td>
<td>0.018 (0.001)</td>
<td>2.56</td>
<td>6.3 (0.3)</td>
<td>17.4 (0.6)</td>
</tr>
<tr>
<td>B</td>
<td>0.273 (0.014)</td>
<td>0.486 (0.040)</td>
<td>0.241 (0.040)</td>
<td>0.017 (0.001)</td>
<td>2.62</td>
<td>7.1 (0.2)</td>
<td>17.8 (0.7)</td>
</tr>
<tr>
<td>C</td>
<td>0.275 (0.013)</td>
<td>0.464 (0.019)</td>
<td>0.261 (0.026)</td>
<td>0.015 (0.001)</td>
<td>2.61</td>
<td>7.2 (0.2)</td>
<td>17.6 (0.5)</td>
</tr>
<tr>
<td>0.3–0.5-m depth</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>0.307 (0.018)</td>
<td>0.487 (0.017)</td>
<td>0.206 (0.030)</td>
<td>0.008 (0.000)</td>
<td>2.60</td>
<td>6.6 (0.3)</td>
<td>18.0 (0.6)</td>
</tr>
<tr>
<td>C</td>
<td>0.269 (0.038)</td>
<td>0.450 (0.026)</td>
<td>0.281 (0.054)</td>
<td>0.007 (0.004)</td>
<td>2.65</td>
<td>7.4 (0.2)</td>
<td>15.3 (2.6)</td>
</tr>
</tbody>
</table>

† Means and standard deviations (in parentheses) of 12 (topsoil) and eight (subsoil) sampling locations.

Table 2. Soil physical and biological properties prior to compaction, from samples taken during October 2013, i.e., 5 mo before the compaction event, including bulk density (BD), total porosity (TP), air-filled porosity at 100 hPa water suction (ɛ₉₀₋₁₀₀hPa), relative gas diffusion coefficient at 100 hPa water suction (Dₑ₋₁₀₀hPa / D₀), air permeability at 100 hPa water suction (kₑ₋₁₀₀hPa), saturated hydraulic conductivity (Kₛₑₑ), microbial biomass (MB, as microbial C [Cₘᵦₑ]), and basal respiration (BR).

<table>
<thead>
<tr>
<th>Depth</th>
<th>BD (Mg m⁻³)</th>
<th>TP (m³ m⁻³)</th>
<th>ɛₑ₋₁₀₀hPa</th>
<th>Dₑ₋₁₀₀hPa / D₀</th>
<th>log kₑ₋₁₀₀hPa</th>
<th>log Kₛₑₑ</th>
<th>MB (mg Cₘᵦₑ kg⁻¹)</th>
<th>BR (μg CO₂-C g⁻¹ h⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>m</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.1</td>
<td>1.38 (0.01)†</td>
<td>0.47 (0.01)</td>
<td>0.09 (0.01)</td>
<td>0.019 (0.002)</td>
<td>1.29 (0.15)</td>
<td>2.08 (0.07)</td>
<td>482.5 (15.4)</td>
<td>0.55 (0.05)</td>
</tr>
<tr>
<td>0.3</td>
<td>1.52 (0.01)</td>
<td>0.42 (0.01)</td>
<td>0.06 (0.01)</td>
<td>0.010 (0.001)</td>
<td>0.93 (0.02)</td>
<td>2.18 (0.20)</td>
<td>161.8 (0.81)</td>
<td>0.33 (0.10)</td>
</tr>
<tr>
<td>0.6</td>
<td>1.53 (0.03)</td>
<td>0.42 (0.01)</td>
<td>0.04 (0.01)</td>
<td>0.005 (0.001)</td>
<td>0.43 (0.07)</td>
<td>1.63 (0.10)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

† Means and standard errors (in parentheses; n = 3 experimental blocks).
† 0–0.2-m depth.
§ 0.3–0.5-m depth.
avoid these zones. On the other hand, it could be argued that recovery in CWT would be faster than in CEP because the compacted zone has a larger interface with uncompacted soil, from which recovery could be initiated. Treatment CWT would more often occur in practice; however, modern agricultural machinery (e.g., self-propelled sugarbeet harvesters, self-propelled forage harvesters, self-propelled slurry tankers) are equipped with offset steering and therefore track the entire surface area as in treatment CEP.

**Post-compaction Soil Management and Cropping Systems**

The SSO includes four post-compaction soil management–cropping systems: bare soil (BS), permanent grass (PG), crop rotation under no-till (NT), and crop rotation under conventional tillage (CT). No farming traffic and no mechanical disturbance are allowed in systems BS and PG; these systems therefore provide insights into uninterrupted natural processes of soil structure regeneration, with plants signifying normal biological activity (PG) and without plants suggesting reduced biological activity (BS). The two cropping systems (NT and CT) enable insights into soil structure recovery under common agricultural practices with minimal (NT) and conventional mechanical soil disturbance (CT).

Plants on the bare soil treatments (BS) are suppressed by periodic application of nonselective herbicides (glyphosate [N-(phosphonomethyl)glycine], glufosinate-ammonium [2-amino-4-(hydroxymethylphosphinyl)butanoic acid monoammonium salt]). The herbicide is applied manually using a knapsack sprayer with a 2.5-m swath width. The treatment PG represents a grass mixture that was established in spring 2013 (see above), i.e., the grass survived the compaction without any resowing. The grass is cut four to five times per year using a small self-propelled motor mower (total mass $\sim 200$ kg). The cut grass is then manually removed from the experimental area.

Triticale ($\times$Triticosecale, cultivar Trado) was sown in the NT and CT plots on 3 Apr. 2014, i.e., 8 d after compaction. Glyphosate was applied to the BS and NT plots on 29 Mar. 2014 to control the ley that was established in 2013 (see above). The crop was sown without any prior tillage with a no-till drill in the NT plots, while the soil was moldboard plowed to about 0.25 m and harrowed to about the 0.06-m depth using a rotavator on 2 Apr. 2014 in the CT plots. Fertilization of the NT and CT plots is performed according to the Swiss fertilization recommendation (GRUDAF) (Flisch et al., 2009), and crop protection (weed and disease control) in NT and CT is done according to the principles of "integrated pest management" aiming to suppress pest populations below the economic injury level, largely relying on pesticides. The crop rotation for NT and CT, to date, is: triticale ($\times$Triticosecale, 2014)–silage maize ($Zea$ mays L., 2015)–winter wheat ($Triticum$ aestivum L., 2016)–winter rapeseed ($Brassica$ napus L., 2017).

**Monitoring and Sampling Concept**

Observations at the SSO include continuous monitoring of soil state variables (soil moisture, temperature, soil air CO$_2$ and O$_2$...
Soil sampling and in situ measurements during pre-compaction soil characterization were performed at 12 randomly selected sampling or measuring locations per experimental block. Earthworm sampling was done at five randomly selected locations within each block. Geophysical measurements were performed in transects, with one electrical resistivity tomography (ERT) transect (48 electrodes, 1-m spacing) per block and three ground-penetrating radar (GPR) transects per block (one transect in the north–south direction and two in the east–west direction). Selected soil properties measured during the pre-compaction soil characterization are given in Table 2. The ERT and GPR results revealed only subtle lateral heterogeneities within and between transects, thereby indicating similar soil profiles across the SSO.

Post-compaction soil characterization was done 2 wk after compaction except for earthworm sampling, which was performed 2 mo after compaction. Three sampling or measuring locations were randomly selected in each NOC and CEP plot. For CWT, sampling or measuring was done at three positions, namely the track center, track edge, and between tracks, with three randomly selected sampling or measuring locations per plot for each position, amounting to a total of nine sampling or measuring locations in each CWT plot. Earthworms were sampled at two randomly selected locations per experimental block in the CEP and NOC plots.

Sampling and measurements during post-compaction monitoring will be performed at intervals of half a year (within the first 1–2 yr after compaction) to 1 yr (from the third year after compaction; and possibly larger time intervals at later stages). The sampling and measuring locations will be randomly selected within each plot. A minimum distance between plot edge and sampling and measuring locations of 1 m, as well as a minimum distance between previous and subsequent sampling and measuring locations of 1 m will be considered. The post-compaction monitoring includes fewer sampling or measuring locations than the post-compaction characterization and may not always include all sampling depths because recovery is expected to be slower at greater depths. Additional measurements such as greenhouse gas emissions could potentially be added later or for some periods of time.

Compaction Stresses in the Soil

During the compaction event, we measured soil mechanical stresses at the 0.2-, 0.4-, and 0.6-m depth using custom-made Bolling probes (Bolling, 1987), which are fluid-inclusion-type probes used to determine mean normal stress (Berli et al., 2006). These measurements were performed between the three experimental blocks (Fig. 2) to avoid damaging the experimental plots. The Bolling probes were installed as described by Keller et al. (2016). We acquired information from four field replicates, i.e., we installed the probes at four different locations within the field. We collected undisturbed soil samples (0.1-m diameter, 0.06-m height) at depths of 0.2, 0.4, and 0.6 m. Sampling locations were adjacent to the locations of soil stress measurements for determination of Poisson’s ratio, ν, in the laboratory according to the method of Eggers et al. (2006) based on uniaxial confined and unconfined compression tests. The mean normal stress, $\sigma_n$, was then calculated from the Bolling probe pressure, $P_i$, and $\nu$ using Eq. [20] and [23] of Berli et al. (2006).

Monitoring Changes in Soil Surface Elevation Using Lidar

As part of the monitoring, we examined the potential of a modern lidar system for capturing soil surface elevation changes due to the prescribed compaction treatment and subsequent rebound. Such a system requires reference points of known position and elevation, so that measurements can be translated to surface elevation changes. Six ground screws (0.8 m in length; Krinner GmbH), serving as fixed points, were drilled into the soil at positions between the experimental plots. During measurements (scanning), spherical targets mounted on ~1.5-m-long aluminum rods are fixed to the ground screws, defining a fixed coordinate system (see also Friedli et al., 2016). Measurements were performed using a Faro Focus 3D S 120 laser scanner (Faro Technologies Inc.) that was mounted on a tripod at a height of about 3.5 m (Friedli et al., 2016). See Friedli et al. (2016) for more details on the scanner and the field setup.

An additional setup was used to perform scans immediately before compaction, immediately after trafficking, and 0.5, 1, and 2 h after trafficking, to quantify vertical surface displacements and potential rebound effects. Wooden square reference plates of 0.25 m$^2$ were placed into the center of two experimental plots (one plate per plot) after the vegetation was cut. Three spheres of 0.1-m diameter mounted on poles at a height of 0.75 m were arranged in a triangle and served as reference points to determine the displacement of the plates. The scanner was mounted on a tripod at a height of 3.5 m above the land surface. The horizontal distance from the scanner to the plates was 12.5 m, and the distance to the spheres was 1 m (one sphere) and 25 m (two far-off spheres). Scans were performed with a measuring rate of 244,000 scan points per second at half of the maximum resolution provided by the system. The distances between single scan points on the plates and the far-off spheres were 3.9 and 7.7 mm, respectively.
In Situ Sensors for Monitoring Physicochemical Soil State Variables

Sensor probe banks (Fig. 2) consisting of probes at various depths (Fig. 3) were installed post-compaction for continuous monitoring of the evolution of soil moisture states, soil temperature, soil air composition (O₂ and CO₂ concentrations), O₂ diffusion rates, and redox states. Installation started 5 d after the compaction event and was completed 10 d after the compaction event. Two banks were established in the CWT plots (Fig. 2): one in the center and one at the edge of a wheel track. The depths considered were 0.1, 0.2, and 0.4 m (all probes), and additionally 0.7 and 1 m (soil moisture probes). The probes were installed either horizontally from a pit or from above ground (Fig. 3). The distance between plot edge and sensing head was at least 0.6 m. In each bank, two to four replicate probes were installed per depth. Probes at the 0.1- and 0.2-m depths in the CT plots are removed and reinstalled at each tillage occasion. The sensor system was automated using CR800 and CR1000 dataloggers and AM16/32B multiplexers (all from Campbell Scientific Ltd.). Wired connection of the sensors to the dataloggers and multiplexers started as soon as the sensor probes were in the ground, but completion of the datalogging system took several weeks. Data are recorded every 30 min (for technical reasons, readings of redox potentials and O₂ diffusion rates are taken only every 8 h; see below) and automatically transferred via General Packet Radio Service (GPRS) communication to a database on a web server (WebDAVIS 3.0, Solexperts AG).

The soil moisture conditions are monitored by measuring soil water suction and soil water content. Soil water suction is measured with a combination of tensiometers (in-house production; Klute, 1986) and calibrated heat dissipation sensors (pF-Meter, ecoTech). The latter sensors yield indirect matric potential measurements and were installed due to the limited range of tensiometers in dry soil (tensiometers fail at water suctions >800 hPa). Soil water content is measured using in-house-produced three-prong time-domain reflectometry (TDR) probes (Jones et al., 2002) and frequency-domain reflectometry (FDR) profilers (EnviroScan, Senteck). The FDR profilers are easily removed from the soil, an operation that needs to be done in the two crop rotation treatments (CT and NT) during field operations (soil tillage, seeding). Some plots are equipped with both TDR and FDR sensors to allow cross comparisons. Two FDR profilers and two tensiometers per depth were installed in Block B at the end of summer 2013 (i.e., more than half a year before the compaction event). Apart from getting a “general feeling” for the soil water dynamics of the site, measurements from these probes were used to define an appropriate time for the compaction event (e.g., soil water content similar to the soil water content at the maximum Proctor density; ASTM, 1992; Aragón et al., 2000; Botta et al., 2008). Soil temperature is measured using standard copper-constantan thermocouples. Soil temperature dynamics affect biological activity and may also carry a structural signature (e.g., higher thermal conductivity for compacted layers).

Redox potentials (E_H) and O₂ diffusion rates (ODR) are monitored with a system consisting of Pt glass electrodes, calomel reference electrodes, and a brass anode, as detailed by Reiser et al. (2012). An 8-h logging interval, from one E_H–ODR data pair to the next, allows relaxation of the electrode polarization and regeneration of the depleted O₂ concentration near the Pt electrode.

Soil O₂ and CO₂ concentrations were initially measured manually at biweekly intervals according to a method described by Weisakopf et al. (2010). Porous polypropylene tubes of 0.5-m length were horizontally installed at the 0.1-, 0.2-, and 0.4-m depths for measurements of O₂ and CO₂ concentrations in the soil air using a CheckMate 9900 head space gas analyzer (PBI Dan-sensor A/S). Probes for continuous measurements were developed later.

Electrical Resistivity Tomography Monitoring and Ground Penetrating Radar

Repeated (two to three times per year) ERT measurements are performed in the SSO in an attempt to observe geophysical signatures of structural changes with time at the plot scale. Stainless steel electrodes were installed along transects (48 electrodes per transect; electrode spacing: 1 m) in the BS and PG plots, as indicated in Fig. 2. The electrodes were left in the ground to ensure that subsequent data acquisitions were acquired with exactly the same electrode geometry. This enables more consistent interpretations and makes it possible to use advanced time-lapse inversion strategies. The ERT data are acquired using a Syscal Switch Pro system (http://www.iris-instruments.com) in a Wenner-Schlumberger electrode configuration. The ERT data are inverted using the algorithm described by Günther et al. (2006) and implemented in the BERT software. Ground-penetrating radar measurements are performed using dual-frequency antennas operating at 300 and 800 MHz (UtilityScan DF by GSSI). The GPR data are analyzed using Reflexw (Sandmeier geophysical research) and in-house MATLAB scripts.

Soil Geochemical Properties

Soil samples are taken from the 0.0- to 0.2- and 0.3- to 0.5-m depth intervals. Soil texture is measured using the pipette method. Soil organic C (SOC) is determined by the wet combustion technique following ISO 10694. Soil pH was measured in CaCl₂. Total N is measured using the Dumas combustion method with an element analyzer (varioMAX CNAnalyser, Elementar Analysensysteme GmbH) (ISO 13878). The concentrations of exchangeable bases, i.e., Ca, K, Mg, Na, and P, are measured by the NH₄OAc method at pH 4.65. All analyses are performed according to Swiss standard protocols (Swiss Federal Research Stations, 1996).

Microbial Biomass

Soil samples are collected at two different depths, 0.0 to 0.2 and 0.3 to 0.5 m. Soil microbial biomass is obtained by two methods: as C_mic estimated according to Anderson and Domsch (1978) and as C_mic−SIR calculated from soil initial respiration (SIR) rates.
Fig. 3. (a) Schematic overview of a sensor bank, showing number and depths (0.1, 0.2, 0.4, 0.7, and 1.0 m) of the various sensor probes (but not necessarily the exact locations) installed from a profile wall at the plot edge through drilled holes of 0.6-m length; and (b) photo of a sensor bank where soil probes have been installed into the profile wall at various depths. The profile wall is at the plot edge.
1986). Air permeability is obtained by measuring the air flow through the sample at an overpressure of 2 hPa, and gas diffusivity is measured in a one-chamber apparatus that uses O2 as the diffusing gas assuming steady-state diffusion (Martínez et al., 2016). A subset of the soil core samples is scanned at 100 hPa water suction with X-ray micro-computed tomography (Phoenix Votmex x s 240, GE Sensing and Inspection Technologies GmbH) for visualization and characterization of soil pore architecture. The soil cores are dried in an oven at 105°C for at least 48 h after the various measurements, and the water content and bulk density are calculated.

Penetration Resistance and Water Infiltration
Penetrometer resistance is measured by means of a hand-pushed Eijkelkamp cone penetrometer (cone base area 1 cm², cone apex angle 60°) to a depth of 0.8 m. Two insertions are made at each sampling location. Steady-state infiltration is measured at the soil surface using a disk permeameter (diameter: 0.2 m) (Perroux and White, 1988). Two infiltration measurements per sampling location are made. The infiltration rates are derived from the cumulative infiltration vs. time relationship.

Earthworm Abundance and Biomass
Earthworms are sampled in an area of 0.5 by 0.5 m using the following procedure. The top ~0.3 m of soil is excavated, and earthworms are collected by hand sorting. Then a 0.5% formaldehyde solution is applied to repel and thus extract earthworms from the subsoil (Kramer et al., 2008). Adults are determined to the species level, juveniles to ecological groups. Abundance and biomass are determined for each taxonomic group.

Crop Response Measurements
Both below- and aboveground crop properties are measured periodically. The root mass depth distribution for treatments PG, NT, and CT is obtained from sampling soil cores (0–0.75-m depth) and washing out roots (Colombi and Walter, 2016). Root architectural traits and root anatomy are measured for the NT and CT treatments as described by Colombi and Walter (2016). Aboveground measures include biomass in the PG plots (separately for grass and legumes) for the first and last cut of a season and plant height, leaf area index, and shoot dry biomass at different stages in NT and CT plots (Colombi and Walter, 2016). Crop productivity on a per-hectare basis, being either grain or silage yield, is determined by harvesting multiple rows of a defined length within the plots (NT and CT).

Initial Compaction Effects
Soil compaction was performed on the ley on 26 Mar. 2014 at an average water content close to field capacity using a self-propelled two-axle fully loaded agricultural self-propelled vehicle. The vehicle was equipped with 1050/50R32 tires inflated to 330 kPa. The wheel load, measured with portable vehicle scales (Radlastwaage WL 103, Haenni Instruments AG) on site, was 8.9 Mg on the front axle and 7.2 Mg on the rear axle (total vehicle mass: 32.2 Mg). Three vehicle passes (forward each time) were conducted on both CEP and CWT plots. The vehicle driving speed was maintained constant at 2 m s⁻¹.

Soil Moisture at Compaction
The soil moisture conditions on the day of compaction (26 Mar. 2014) were “ideal” for effective soil compaction (i.e., the soil water content was close to the water content at the maximum Proctor density; e.g., Aragón et al., 2000; Botta et al., 2008). We measured average soil water suctions, obtained from tensiometers, of 54, 60, 86, and 105 hPa at the 0.1-, 0.2-, 0.4-, and 0.7-m depths, respectively. For the same depths, the volumetric water contents, \( \theta \), obtained from FDR profilers, were 0.35, 0.32, 0.28 and 0.24 m³ m⁻³, respectively, corresponding to gravimetric water contents, \( w \), of 0.26, 0.22, 0.18 and 0.15 g g⁻¹, respectively. These values are slightly above the “optimum water content,” \( w_{opt} \), associated with the maximum density obtained in a Proctor test: Naderi-Boldaji et al. (2016) obtained \( w_{opt} = 0.18 \) g g⁻¹ from soil sampled at the 0.25-m depth in the immediate vicinity of the SSO (on the same field). This is identical to \( w \) measured at 0.4 m but slightly lower than \( w = 0.22 \) g g⁻¹ that we measured at 0.2 m. Note that the optimum water content would be larger than the 0.18 g g⁻¹ reported by Naderi-Boldaji et al. (2016) at the 0.1-m depth because of the higher SOC concentration in the topsoil, and lower than 0.18 g g⁻¹ at 0.7 m because of the lower SOC concentration in the subsoil than at the 0.25-m depth. Hence, \( w \) at the time of the compaction event was close to \( w_{opt} \) at all considered depths.

Soil Stress, Surface Displacement, and Bulk Density
The mean normal stress under the agricultural vehicle decreased from ~150 kPa at 0.2 m to ~50 kPa at 0.5 m (Fig. 4a). The increase in bulk density (Fig. 4b) was large at 0.1 m (from ~1.1 to 1.5 Mg m⁻³) and 0.3 m (from ~1.5 to 1.6 Mg m⁻³), while there was only a small increase in bulk density at the 0.6-m depth. The depth of...
compaction of about half a meter is consistent with the results presented by Keller et al. (2012), who analyzed data from a large number of trafficking experiments on various soil textures and found that vertical strain was observed when the vertical stress exceeded $\sim 40$ kPa. We calculated the vertical soil displacement from the measured changes in bulk density by interpolating between measuring depths, which resulted in a vertical displacement at the soil surface, i.e., rut depth, of 50 mm (Fig. 4c). This compared well with manual measurements using a ruler, from which we obtained rut depths between 37.5 (between tire lugs) and 61.3 mm (on tire lugs). The vertical surface displacement obtained from the lidar measurements (see above) was on average 22 mm. The smaller surface displacement is explained by the fact that the reference plates were lying on the highest points of the soil surface and thus measured the minimum vertical displacement. Lidar measurements at 0.5, 1, and 2 h after compaction did not reveal any rebound.

We estimated the maximum Proctor density, $\rho_{\text{Proctor}}$, using Eq. [8] of Naderi-Boldaji et al. (2016) and the soil textural data given in Table 1 to 1.66 and 1.70 Mg m$^{-3}$ for the 0.1- and 0.3-m depths, respectively. The degree of compactness, DC, given as the ratio of the actual bulk density to $\rho_{\text{Proctor}}$, increased due to compaction from 78 to 90% at the 0.1-m depth, and from 88 to 94% at the 0.3-m depth. Critical limits of DC with respect to crop yield reported in the literature are 84 to 87% (Carter, 1990; Håkansson and Lipiec, 2000; Naderi-Boldaji and Keller, 2016). Using the relationship between DC and the Dexter (2004) soil quality index $S$ developed by Naderi-Boldaji and Keller (2016), the DC values of the 0.1-m depth (topsoil) translate into $S$ values of 0.044 (uncompacted) and 0.027 (compacted), indicating a shift from “good” soil physical quality to “poor” soil physical quality. These estimations suggest that the compaction inflicted in the SSO created soil physical conditions that are considered poor and expected to be limiting for crop development.

### Soil Structure, Transport Properties, and Mechanical Impedance

The internal soil deformation and increase in bulk density (i.e., decrease in total porosity; see above) due to the trafficking by the agricultural vehicle caused significant changes to the soil pore structure and soil transport properties (Table 3). Figure 5 presents...
examples of the pore system (pores detectable on micro-computed tomography images, i.e., pore diameter > 120 μm) of compacted and uncompacted soil at the 0.1- and 0.3-m depths. The decrease in pore connectivity and in porosity is clearly visible at both depths, as also confirmed from measurements of air-filled porosity and gas transport properties (Fig. 6).

The decrease in porosity (Fig. 4b, 5, and 6a) drastically reduced the saturated hydraulic conductivity, $K_{sat}$ (Fig. 6b), air permeability, $k_a$ (Fig. 6c), and the relative gas diffusion coefficient, $D_p/D_0$ (Fig. 6d). The three different air-filled porosities per treatment and depth in Fig. 6c and 6d reflect measurements obtained at three water suction values (30, 100, and 300 hPa, respectively). A significant decrease in air-filled porosity and transport properties was measured at the 0.6-m depth (Fig. 6), although there was only a small increase in bulk density (Fig. 4b). This shows that soil functions related to pore size distribution and pore connectivity are more sensitive to compaction than macroscopic properties such as bulk density (Horn, 2003). Compaction did not affect the relationship between $D_p/D_0$ and $\varepsilon_a$ (Fig. 6d), i.e., it did not affect the specific diffusivity ($\epsilon_D = D_p/D_0 \times \varepsilon_a^{-1}$), but $D_p/D_0$ was substantially decreased in the compacted soil due to lower $\varepsilon_a$. The slope of the log $k_a$ vs. log $\varepsilon_a$ relationship was slightly smaller for compacted soil (Fig. 6c), which would imply a slightly higher specific air permeability ($\epsilon_A = k_a \times \varepsilon_a^{-1}$) at small $\varepsilon_a$ but a slightly lower $\epsilon_A$ at high $\varepsilon_a$. During recovery, it may be expected that $\epsilon_A$ and $\epsilon_D$ initially increase and later decrease again (Jarvis, 2007; Mossadeghi-Björklund et al., 2016).

Water infiltration at the soil surface, measured by means of a disk permeameter, exhibited a more pronounced reduction due to compaction than $K_{sat}$ in the soil profile. Infiltration rates decreased from 8 to 0.001 mm min$^{-1}$, i.e., a decrease of almost three orders of magnitude. In other words, immediately following compaction, the soil surface became nearly impervious, which could be attributed to the excessive smearing of the soil surface by the
Fig. 6. Initial compaction effects on (a) air-filled porosity, $\varepsilon_a$, at different water suctions (circles: uncompacted; squares: compacted; red: 0.1-m depth, blue: 0.3-m depth, green: 0.6-m depth); (b) saturated hydraulic conductivity, $K_{sat}$ (circles: uncompacted; squares: compacted); (c) air permeability, $k_a$, plotted as $\log k_a$ vs. $\log \varepsilon_a$ (circles: uncompacted; squares: compacted; red: 0.1-m depth, blue: 0.3-m depth, green: 0.6-m depth); and (d) relative gas diffusion coefficient, $D_i/D_0$, plotted as a function of $\varepsilon_a$ (circles: uncompacted; squares: compacted; red: 0.1-m depth, blue: 0.3-m depth, green: 0.6-m depth). Note that the legends of Fig. 6d also apply to Fig. 6a and 6c. The data were obtained from soil cores sampled 2 wk after the compaction event. The data represent average values from the three treatments without tillage (i.e., bare soil, permanent grass, and no-till).
agricultural vehicle in addition to the effects of soil compaction. As a consequence, water ponded after rainfall in the compacted plots and in the wheel tracks. We measured an appreciable increase in mechanical resistance in the compacted topsoil (0–0.3-m depth) relative to the uncompacted soil. Penetration resistance was almost 2.5 MPa in the compacted soil at the 0.1- to 0.25-m depth, while the mechanical resistance at this depth was ~1.3 MPa in the uncompacted soil. Root elongation rates decrease with increasing soil penetration resistance (e.g., Dexter, 1987). A resistance of 2 MPa is often considered critical for root growth (Taylor et al., 1966; da Silva et al., 1994); it may correspond to a value where root elongation rates are about half of the maximum root elongation rate (Dexter, 1987).

Earthworms and Microbial Biomass
The earthworm population was drastically reduced due to compaction. The estimated biomass in the compacted plots was only 30% (epigeics) to 41% (anecics) of that in the uncompacted soil (Table 4). The decrease was probably caused by increased mortality rates during the compaction event (i.e., during loading; McKenzie et al., 2009), as well as less favorable living conditions after compaction. There were no immediate effects of soil compaction on microbial biomass and respiration. However, we expect microbial communities and hence microbial biomass and respiration to adapt to the changed soil physical conditions with time (Hartmann et al., 2013).

Electrical Resistivity Tomography
An example of a preliminary analysis is given for the bare soil in Block B (Fig. 7). In this figure, the relative difference between the estimated electrical resistivity on 7 Apr. 2014 (12 d after compaction) and 21 Mar. 2014 (before the compaction) is shown. The soil electrical resistivity decreased by 10 to 15% for the part of the block that had undergone CEP treatment, and decreases were also found under the wheel tracks for the CWT treatment. These results were obtained by an inversion process that smoothens actual variations in resistivity, which implies that the compaction-affected depth range is probably smaller than the approximately 0 to 1 m that is indicated in this figure.

<table>
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<th>Ecophysiological category</th>
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<th>2014</th>
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<td>Epigeic</td>
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</tr>
<tr>
<td>Endogeic</td>
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<td>Anecic</td>
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<tr>
<td>Total earthworm biomass</td>
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Outlook
The SSO aims at providing long-term data on the post-compaction soil structure evolution under contrasting post-compaction soil management regimes. The long-term time scale and tight intervals of observations (continuous measurements provided by various sensor probes in combination with regular sampling and in situ measurements) are crucial. We expect that the observations will allow quantification of recovery rates and recovery times of compacted soil and a better understanding of the (relative importance of the) recovery mechanisms, considering both natural biotic and abiotic mechanisms as well as soil tillage. Furthermore, we expect that the generated knowledge will help define strategies and guidelines for accelerating soil structure recovery, and more generally, improving soil structure in modern agriculture.

Sampling and in situ measurements have so far been done at half-yearly intervals after the compaction event, and sensor probes were installed in the weeks after the compaction event. Initial compaction increased the soil bulk density to about 0.5-m depth, decreased soil gas and water transport capability (air permeability, gas diffusivity, saturated hydraulic conductivity), and increased mechanical impedance. Water infiltration at the soil surface was drastically reduced (see also Fig. 8). Initial results from the post-compaction monitoring indicate projected recovery rates of years to decades, with different recovery rates for different properties and decreasing recovery rates with soil depth. Furthermore, the data indicate that soil tillage may immediately recover macroscopic soil total porosity but not soil functions (e.g., gas transport properties).

It would be highly desirable to establish similar SSOs under different soil and climatic conditions based on the concept outlined here. Additional SSOs could also include different compaction levels or different post-compaction soil management regimes. Data from other SSOs would increase our knowledge on how soil compaction recovery (and more generally, soil structure dynamics) is influenced by soil, climate, crops, and their interactions, allow estimates of compaction recovery rates and times across a wide range of soil and climatic conditions, and provide a wider basis for the development of soil management strategies.
Fig. 8. Ponding water after rainfall in (a) compacted plots and (b) wheel tracks of the bare soil treatment in April 2015 (i.e., 1 yr after compaction).


